- 1 Effect of multinutrient insufficiency on markers of one carbon metabolism in young
- 2 women: response to a methionine load
- 3 Running title: Multinutrient insufficiency and 1-C metabolism
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## 28 Abstract

29 Background: Multinutrient insufficiencies as a consequence of nutritional and economic factors

30 are common in India and other developing countries. We have examined the impact of multi-

nutrient insufficiency on markers of one carbon metabolism in the blood, and response to a

32 methionine load in clinically healthy young women.

**Design & Methods:** Young women from Pune, India (n=10) and Cleveland, USA (n=13) were studied. Blood samples were obtained in the basal state and following an oral methionine load (50mg/kg of body weight in orange juice). Plasma concentrations of vitamin B12, folate and B6 were measured in the basal state. The effect of methionine load on the levels of methionine, total homocysteine, cysteine, glutathione and amino acids was examined.

**Results:** Indian women were significantly shorter and lighter compared with the American women and had lower plasma concentration of vitamins B12, folate and B6, essential amino acids and glutathione, but higher concentration of total homocysteine. The homocysteine response to methionine load was higher in Indian women. The plasma concentrations of glycine and serine increased in the Indian women after methionine (in juice) load. A significant negative correlation between plasma B6 and homocysteine (r= -0.70), and plasma folate and glycine and serine levels were observed in the Indian group (P<0.05) but not in the American group.

45 Conclusion: Multi-nutrient insufficiency in the Indian women caused unique changes in markers
46 of whole body protein and one carbon metabolism. These data would be useful in developing
47 nutrient intervention strategies.

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#### 55 Introduction

Folate one-carbon transfers, present ubiquitously in every cell in the body, are key components of 56 cell metabolism. They are involved in transfer of methyl groups for biological methylation 57 reactions including synthesis of nucleotides. In addition to folate, vitamin B12 (B12) and vitamin 58 B6 (B6) along with insulin and glucagon are respectively key co-factors and hormonal regulators 59 of one carbon metabolism in-vivo. Perturbations of one carbon metabolism as a consequence of 60 changes in nutrient status of the individual or of hormonal, and environmental interactions, have 61 been related to birth defects, cancer, metabolic disorders, cardio-vascular disease and to aberrant 62 DNA methylation patterns. Because of their critical role in whole body and cell metabolism, a 63 number of intracellular and circulating biomarkers of nutrient deficiencies related to one carbon 64 metabolism have been identified and validated in order to develop and monitor intervention 65 strategies for the 'at risk' populations.<sup>1-5</sup> Studies in both human and animal models have examined 66 the impact of micro-nutrient deficiency or insufficiency on biomarkers of one carbon metabolism 67 in the plasma and tissues.<sup>1,3</sup> Most of these studies have examined the association between a single 68 nutrient deficiency with the identified biochemical marker/s.<sup>1-4</sup> Isolated single nutrient 69 deficiencies although observed in otherwise well-nourished populations, they are uncommon in 70 the undernourished populations, particularly in relation to folate, B12, B6, and protein, the 71 nutrients that impact one carbon metabolism<sup>6,7,8</sup>. 72

The folate-methionine cycle and its key regulatory cofactors are displayed in Figure 1. As shown, serine and glycine are the major contributors of one carbon (1C) units. In this process, serine is converted reversibly to glycine in a B6 dependent reaction catalysed by serine hydroxymethyl transferase and the 1C unit is transferred to tetrahydrofolate (THF), to from 5,10-methylene tetrahydrofolate. Glycine contributes 1C units via the glycine cleavage system to 78 tetrahydrofolate. Methionine is activated to form s-adenosylmethionine (SAM) by methionine adinosyl transferase and ATP. SAM is the key methyl donor for methylation reactions catalysed 79 by various methyltransferases and in the process is converted to s-adenosyl homocysteine (SAH) 80 and ultimately to homocysteine. SAM also can be converted, in the liver, to SAH by glycine-n-81 methyl transferase (GNMT). Vitamin A is a transcriptional or translational regulator of GNMT 82 activity. Homocysteine can either be converted back to methionine (remethylation) catalysed by 83 methionine synthase or metabolized to cystathionine and cysteine (transsulfuration). B12 is the 84 cofactor for methionine synthase (5-methyltetrahydrofolate homocysteine methyltransferase) 85 responsible for the transfer of methyl group of 5-methyl tetrahydrofolate to homocysteine to 86 form methionine (remethylation). The two enzymes of the transsulfuration cascade require B6 as 87 a cofactor. Isolated deficiency of the cofactors (B6, B12 or folate) can result in increased levels 88 of the precursor and lower levels of the immediate product. In addition, isocaloric protein 89 restriction in animal models and lower dietary protein intake in humans has been shown to result 90 in increase in plasma levels of homocysteine, serine and glycine<sup>9-11</sup>. The combined effect of 91 deficiency or insufficiency of these micronutrients and lower protein intake has not been 92 examined. In the present study, we have examined the integrated changes in one carbon 93 metabolism in response to multi-nutrient insufficiency in an otherwise "healthy" group of Indian 94 women and compared with a group of "nutritionally sufficient" American women. 95

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#### 97 Subjects and Methods

98 The study participants in India (n=10) were young healthy female staff members of the King 99 Edward Memorial Hospital Research Centre, Pune. Healthy young women in Cleveland, USA 100 (n=13) were recruited by advertisement. The study protocol was approved by the Ethics 101 Committee of the KEM Hospital Research Centre and by the Institutional Review Board of the 102 Cleveland Clinic. Written informed consent was obtained from the participants after fully 103 explaining the procedure.

The subjects reported to the research unit at 7 am following a 12 hour fast. Height and weight 104 were measured as per the standard protocol. An indwelling cannula was placed in an antecubital 105 vein and subjects were allowed to rest for half an hour. After obtaining a basal blood sample, L-106 methionine (50 mg/kg body weight) was given in orange juice along with a standardized low 107 methionine breakfast (estimated total methionine content ~58mg). We elected to give a lower 108 109 dose of methionine instead of the usual 100mg/kg body weight, and perform a short 5 hour instead of 6 hour test for the following reasons: (i) Results of the dose response studies show that 110 50mg/kg can give data similar to the higher dose without compromising the sensitivity of the 111 test<sup>12,13</sup> (ii) a short 3 hour test was as good as the standard 6 hour test in identifying subjects with 112 hyperhomocysteinemia<sup>14,15</sup>, and (iii) our ultimate goal is to do these studies during pregnancy 113 and we were concerned about the potential toxicity of methionine and homocysteine with the 114 higher dose to the mother and the developing conceptus. Blood samples, in EDTA tubes, were 115 drawn at hourly interval for the next 5 hours. Five subjects in the Cleveland group and five in the 116 117 Indian group received methionine load without the accompanying breakfast. Since there was no significant difference in the response to methionine load in those with or without breakfast the 118 data were combined. Blood samples were centrifuged in cold and the plasma was stored at -80°C 119 120 for analysis later.

# 121 Laboratory analysis

Amino acid concentration in plasma were measured by HPLC using an OPA derivative and a fluorescence detector as described.<sup>16</sup> Total homocysteine, total cysteine, and glutathione in the plasma were measured using HPLC.<sup>17</sup> Plasma formate levels were measured by an isotopedilution GC-MS method as described by Lammarre et al.<sup>18</sup>

Vitamin B6 (Pyridoxal-5-phosphate and Pyridoxal) was measured using commercially available 126 HPLC kit (RECIPE GmbH, Germany) using post column derivatisation and fluorescence 127 detector. Plasma cobalamin (vitamin B12) and folate were measured by microbiological assay 128 using a colistin sulfate-resistant strain of Lactobacillus leichmanii and a chloramphenicol-129 resistant strain of Lactobacillus casei respectively<sup>19, 20</sup>. The coefficients of variation for B12 and 130 folate measurement in the plasma were < 8% and for B6 it was < 5%. High sensitive C-reactive 131 protein (hsCRP) was measured by high-sensitivity ELISA kit (United Biotech, Mountain View, 132 CA, USA) with inter and intra batch cv <11%. 133

### 134 Statistical Methods

We performed descriptive statistics and checked for normality of the data. Since data were 135 skewed and the sample size small, we used non parametric methods for statistical analysis. Data 136 are presented as median and 25<sup>th</sup> and 75<sup>th</sup> percentiles. Association of homocysteine with B12. 137 folate and B6 was analysed by Spearman's rank correlation coefficient. Differences in various 138 parameters between Indian and US participants were analyzed by Mann Whitney U test. The 139 incremental (above basal) area under the curve for homocysteine and methionine (basal to 5 140 hour) was computed using trapezoidal rule. Multiple linear regression analysis (MLRA) was 141 used to compute variation in outcome variable (homocysteine) explained by exposure variables 142 (B6, B12 and folate). Statistical analyses were performed using SPSS 16 (SPSS Inc. Chicago 143 US). 144

Based on the published literature, the total homocysteine values in Indians is: 23.2 (13.1) and 10.4 (3.6) in western population. Therefore sample size of 8 provide a power of 80% at 5% level of significance and sample size of 11 in each group provides a power of 90% at 5% level of significance. Hence, we have a chosen sample size of 10 in Indians and 13 in US.

# 149 **Results**

Indian women were on average 30 yrs old and predominantly vegetarian. American women were of similar age, and were non-vegetarian in their dietary habits (Table 1). The Indian women were on average 20 cm shorter and 20 kg lighter, though the BMI was not significantly different in the two groups. The plasma levels of B6, folate and B12 in the US group were in the accepted 'reference' range. In contrast, the levels of B6, folate and B12 were significantly lower in the Indian subjects. The plasma levels of hsCRP were not different amongst the two groups.

The plasma levels of amino acids during fasting are displayed in Table 2. As shown, the levels of essential amino acids (valine, phenylalanine, leucine, isoleucine, lysine and methionine) were significantly lower in the Indian women. The levels of histidine and aminobutyric acid also were lower in the Indian women. The concentration of serine and glycine although higher in the Indian group, were not statistically different.

161 In contrast to lower plasma levels of methionine, plasma levels of homocysteine were markedly

higher in the Indian women as compared to those in the American women (Indians: 20.4 (16.4,

163 24.4), Americans: 7.9 (6.8, 8.9) P<0.001). Total plasma glutathione levels (Indians: 3.8 (2.2,

164 3.9), Americans: 6.5 (5.4, 7.5) P<0.001) were lower in the Indian subjects. Total cysteine

165 concentration in the plasma was similar in the two groups. The levels of formate in the plasma

were markedly higher in the Indian women (Indians: 182.9 (167.9, 190.3) micromoles/l,

167 Americans: 39.9(37.2, 48.2) P=0.006).

After the oral methionine load, plasma levels of methionine rose, reaching a peak at one hour and 168 then gradually declined over the next four hours. The magnitude of increase in methionine levels 169 from basal to 1hour was similar in the Indian and American women. As shown in Figure 2, the 170 shape of the methionine curve and the incremental area under the curve were indistinguishable in 171 the two groups. Plasma homocysteine levels rose linearly in both groups. In the American 172 women, it reached a plateau (~  $20\mu$ M/l) by two hours and remained unchanged for the next three 173 hours. In contrast, plasma levels of homocysteine continued to increase in the Indian subjects 174 until 4hours reaching a plateau of  $\sim$ 41µM/l between the 4<sup>th</sup> and the 5<sup>th</sup> hour. 175

176 The rise in homocysteine levels from basal was significantly greater in the Indian subjects at 4 and 5 hours (P<0.01). The incremental area under the curve was not significantly different 177 between the two groups but approached significance following the removal of one outlier (area: 178 179 94.9 µM/l.5 hours) in the US group (Indians: 62.03 (43.9, 77.6), Americans: 49.6 (44.7, 50.8)  $\mu$ M/l.5hours P = 0.06). Plasma levels of total cysteine remained stable in the Indian and 180 American women, following the methionine load. Methionine load caused a small rise in the 181 plasma levels of total glutathione in Indian women but an insignificant increase in the American 182 women (Figure 2). 183

The changes in representative amino acids in the plasma following a methionine load in orange juice are displayed in Figure 3. There was a significant increase in plasma concentration of glycine and serine in the Indian women but not in the nutritionally sufficient American women. The increase in glycine and serine was seen in the Indian women with and without breakfast suggesting that it was due to the carbohydrate load in the orange juice. Plasma concentration of alanine peaked in both groups at 1hr although the magnitude of increase was less in the Indian women. Plasma levels of taurine showed an increase in Indians and a decrease in the Americans. As anticipated the levels of all essential amino acids decreased following nutritional (breakfast and orange juice) load<sup>21</sup>. The data of leucine and phenylalanine are displayed in Figure 3.

Plasma total homocysteine levels during fasting were not correlated with circulating 194 levels of vitamins B6, folate and B12 and methionine in the American women. In contrast, in the 195 Indian women plasma homocysteine was inversely correlated with vitamin B6 levels both in the 196 fasting state and at five hours after methionine load (Figure 4; basal r=-0.68, P<0.05 and 5 hour 197 r = -0.70, P<0.05). The incremental change in homocysteine concentration was significantly 198 correlated with B12 concentration only in the American group (r=-0.73, P<0.01). Multiple linear 199 200 regression analysis showed that circulating levels of vitamins B6, folate and B12 explained 26% of the variance in the plasma basal total homocysteine levels in the American women; in the 201 Indian women this figure was 57%. On the other hand, 82% of the difference in the circulating 202 203 levels of basal total homocysteine in the two groups of women was explained by the difference in the levels of vitamins B6, folate and B12. A significant negative correlation between plasma 204 folate levels and plasma glycine (r= -0.842, P<0.01) and serine (r= -0.697 P<0.05) levels were 205 observed only in the Indian group. There was no correlation between glycine, serine, histidine, 206 methionine and plasma levels of B12 or B6 in either group. 207

208 Discussion

209 Our data show that multi-nutrient deficiencies in the Indian women (vitamins regulating one carbon metabolism) resulted in substantially elevated homocysteine concentrations and 210 lower levels of essential amino acids in the plasma. Oral methionine load showed that the 211 nutritionally compromised Indian women could absorb and dispose off methionine equally 212 efficiently as the nutritionally sufficient American women. However, there was a greater increase 213 in plasma homocysteine concentration in the Indian women. Additionally, there was an increase 214 in the plasma serine and glycine concentration in the Indian women only, likely in response to 215 the carbohydrate load (orange juice) administered with methionine. 216

The present data should be examined in the following context. The dietary habits of 217 Indians from this region are mostly vegetarian with relative lower quantity and quality of protein 218 and lower dietary source of vitamins.<sup>22</sup> This was reflected in the markedly low levels of B12 in 219 the Indian women in this study, which has been previously reported in the vegetarians. <sup>22-25</sup> The 220 221 plasma levels of folate and B6 also were lower in the Indian women. In contrast the American women were all non-vegetarian with higher daily intake of dietary protein. In addition, the folate 222 intake of American subjects was higher due to the mandatory fortification of flour.<sup>26</sup> Given the 223 critical role in one carbon transfers, and as cofactors at specific steps in folate and methionine 224 metabolism, the inadequate intake of these nutrients individually will result in unique changes in 225 one carbon metabolism and the circulating levels of related biochemicals (Figure 1). However, 226 the combined effect of simultaneous insufficiency of these nutrients could be different due to the 227 opposing effect of some of them. For example, lower protein intake results in higher rate of 228 transmethylation of methionine while folate and B12 insufficiency causes a lower rate of 229 methylation of homocysteine. The combined effect of these nutrient insufficiencies has not been 230

examined in humans. The present data reports the net effect of the insufficiency of thesenutrients on one carbon metabolism.

The concentrations of essential amino acids in the plasma were significantly lower in the 233 Indian women in the fasting state. Since breakdown of proteins in the body, primarily skeletal 234 muscle, is the major source of essential amino acids in the plasma, our data suggest a lower rate 235 of protein breakdown or protein turnover in the Indian women. Although the present data cannot 236 delineate the cause of lower rate of protein turnover in these subjects, it is likely to be related to 237 lower dietary intake of proteins and consequent attempt at conservation of nitrogen. Dietary 238 restriction of protein in healthy humans has been shown to cause a decrease in whole-body 239 proteolysis as measured by the rate of appearance of leucine and a decrease in the rate of 240 oxidation of leucine/protein<sup>27, 28</sup> and cause a decrease in the rate of oxidation of leucine in the 241 rat.<sup>29</sup> These changes in essential amino acids were associated with small increase in the levels of 242 243 glycine and serine in the plasma. The latter has been shown to increase in humans and in laboratory animals when dietary proteins are restricted.<sup>30-33</sup> Tracer isotope studies have shown 244 that increase in serine and glycine were the consequence of increased rates of de-novo synthesis 245 of these amino acids <sup>9,34</sup>. The higher levels of glycine and serine and their increased rates of 246 synthesis may be related to the hepatic induction of PPARa as a result of low protein intake.<sup>9</sup> 247 Data in literature show that administration of PPARa agonist in mice results in increased levels 248 and rate of turnover of glycine and serine in the plasma.<sup>35</sup> The physiological significance of 249 changes in glycine and serine metabolism during protein restriction, other than as source of 250 methyl groups, has not been determined. It has been postulated that restriction of dietary protein 251 results in higher methylation demand and a high rate transmethylation and consequently high 252 rate of synthesis of serine and glycine.<sup>9</sup> The negative correlation between plasma levels of folate 253

and plasma levels of glycine and serine during fasting in the Indian women suggests that in addition to low protein, lower folate also may contribute to the higher levels of serine and glycine by attenuating the folate cycle.

A decrease in essential amino acids levels in the plasma was seen in all subjects 257 following the administration of methionine mixed with orange juice and a low methionine 258 breakfast, likely due to the expected suppression of whole body breakdown of proteins in 259 response to carbohydrate (juice) and nutrients (breakfast) and associated increase in insulin.<sup>36,21</sup> 260 In contrast there was an increase in plasma concentration of glycine and serine in the 261 nutritionally insufficient Indian women (Figure 3). These data suggest an active pathway for the 262 synthesis of serine in the liver induced by low protein intake and rapid conversion of dietary 263 carbohydrates into serine and glycine.<sup>37</sup> 264

The fasting plasma tHcy was markedly higher and that of glutathione was lower in the 265 Indian women. There was a significant negative correlation between plasma levels of tHcv and 266 267 pyridoxal phosphate levels (Figure 4), suggesting a dominant contribution of lower rate of transsulfuration to tHcy levels. The plasma levels of tHcy were not related to folate or B12 levels 268 in this small group of women. None the less, the lower folate and B12 levels in the Indian 269 women, by attenuating methylation of homocysteine, also would contribute to the increase in its 270 plasma levels (Figure 1). The higher levels of formate in the plasma, in the Indian women, are 271 consistent with an impaired rate of remethylation of homocysteine in these subjects.<sup>38</sup> The cut off 272 values at which a steep increase in plasma homocysteine concentration occurs in B12 273 insufficiency have been reported to be much higher (200-300pmol/l) than those seen here in the 274 Indian women.<sup>4, 39</sup> In addition lower protein intake by increasing the methionine cycle would 275 also result in higher homocysteine.<sup>9</sup> The individual contribution of inadequacies of these 276

nutrients to the higher levels tHcy cannot be discerned from the present data. These observations underscore the importance of examining the impact of multinutrient deficiencies on metabolic biomarkers and the need for studies using isotopic tracers. The mechanism of lower concentration of glutathione in the plasma is unclear. It was probably not related to lower rate of transsulfuration due to lower B6 levels since the plasma levels of total cysteine were not different in the two groups. The lower levels could be the result of hormonally mediated decrease in glutathione synthesis as a result of altered nutritional state.<sup>40</sup>

We did the methionine load studies in order to (a) describe the net effect of multi-nutrient 284 insufficiencies on one carbon metabolism and (b) to possibly reveal the contribution of 285 transmethylation and transsulfuration of methionine to the observed changes in one carbon 286 metabolism. As shown in figure 2 the plasma methionine response to oral load of 50mg/kg body 287 weight of methionine was similar in Indian and American subjects, both in terms of plasma 288 289 levels of methionine and of calculated incremental area under the curve. These data suggest that there was no difference in the two groups in relation to gastrointestinal absorption, the first pass 290 metabolism and disposal of methionine. In contrast to the disposal of methionine, the 291 292 incremental increase in plasma homocysteine concentrations and the area under the tHcy response curve were different in the two groups. However, it should be underscored that the 293 changes in plasma homocysteine are a net effect of both transsulfuration and remethylation and 294 therefore any differences in these processes cannot be evaluated from these data. Following a 295 methionine load, there would be an increase in the intracellular concentration of s-296 adenosylmethionine (SAM). SAM is an allosteric inhibitor of methylenetetrahydrofolate 297 reductase (MTHFR) and would result in a decrease in the remethylation of homocysteine via 298 methionine synthase.<sup>41-43</sup> SAM is also allosteric activator of cystathionine beta synthase and 299

would cause an increase in transsulfuration pathway.<sup>40,41</sup> Thus the higher plasma tHcy levels in 300 the Indian women after methionine load, in the presence of similar levels of methionine, suggest 301 a decrease in the disposal of homocysteine via the transsulfuration cascade. The significant 302 correlation between plasma B6 levels and the plasma tHcy levels after methionine load support 303 this inference. The mechanism of the observed changes in total glutathione in the plasma i.e. a 304 decrease in the American women and increase in Indian women is not clear. Tracer isotope 305 labelled methionine studies would be required to further interrogate the impact of multiple micro 306 and macro-nutrient insufficiencies on components of methionine metabolism. 307

In summary, we have identified characteristic perturbations in one carbon metabolism and circulating levels of amino acids in response to multi-nutrient deficiency in the Indian women. A significant decrease in concentrations of essential amino acids in the plasma and an increase in serine and glycine suggest a lower isocaloric protein intake. Low B12 and B6 status resulted in higher homocysteine levels in the basal state and a higher homocysteine response to a methionine load. The net impact of these nutritional insufficiencies on transmethylation and transsulfuration of methionine will require careful tracer isotope studies.

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# **References**

| 322 | 1. | Mason J B. Biomarkers of nutrient exposure and status in one-carbon (methyl)                    |
|-----|----|---|
| 323 |    | metabolism. J of Nutrition 2003; 133: 941s-947s.  |
| 324 | 2. | King W D, V Ho, L Dodds, S L Perkins, R I Casson, T E Massey. Relationships among               |
| 325 |    | biomarkers of one-carbon metabolism. Mol Biol Rep 2012; 39: 7805-7812.                          |
| 326 | 3. | Lamers Y. Indicators and methods for folate, vitamin B-12, and vitamin B-6 status               |
| 327 |    | assessment in humans. <i>Current opinion in clinical nutrition and metabolic care</i> 2011; 14: |
| 328 |    | 445-454.  |
| 329 | 4. | Selhub J, PF Jacques, G Dallal, S Choumenkovitch, G Rogers. The use of blood                    |
| 330 |    | concentrations of vitamins and their respective functional indicators to define folate and      |
| 331 |    | vitamin B 12 status. Food and Nutrition bulletin 2008; 29: S67-S73.                             |
| 332 | 5. | Vogiatzoglou A, A Oulhaj, A D Smith, E Nurk, C A Drevon, P M Ueland et al.                      |
| 333 |    | Determinants of plasma methylmalonic acid in a large population: implications for               |
| 334 |    | assessment of vitamin B 12 status. Clinical Chemistry 2009, 55: 2198-2206.                      |
| 335 | 6. | Bhardwaj A, Kumar D, Raina SK, Bansal P, Bhushan S, Chander V. Rapid Assessment                 |
| 336 |    | for Coexistence of Vitamin B12 and Iron Deficiency Anemia among Adolescent Males                |
| 337 |    | and Females in Northern Himalayan State of India. Anemia 2013; 959605.                          |
| 338 | 7. | Food and Nutrition Bulletin Volume 29, Supplement 1, June 2008 Folate and vitamin $B_{12}$      |
| 339 |    | deficiencies: Proceedings of a WHO Technical Consultation held 18-21 October, 2005,             |
| 340 |    | in Geneva, Switzerland  |
| 341 | 8. | Sukla KK Nagar R Raman R. Vitamin-B12 and folate deficiency, major contributing                 |

342 factors for anemia: A population based study. *e-SPEN Journal* 9 (2014) e45-e48

| 343 | 9.  | Kalhan SC, SO Uppal, JL Moorman, C Bennett, LL Gruca, PS Parimi et al. Metabolic          |
|-----|-----|---|
| 344 |     | and genomic response to dietary isocaloric protein restriction in the rat. J Biol Chem    |
| 345 |     | 2011; <b>286</b> : 5266-5277.   |
| 346 | 10. | Ingenbleek Y, E Hardilliera, L Jung. Subclinical protein malnutrition is a determinant of |
| 347 |     | hyperhomocysteinemia. Nutrtion 2002; 18: 40-46.   |
| 348 | 11. | Ingenbleek Y, and KS McCully. Vegetrainism produces subclinical malnutrition,             |
| 349 |     | hyperhomocysteinemia and atherogenesis. Nutrition 2012; 28: 148-153.                      |
| 350 | 12. | Chambers JC, Ueland PM, Wright M, Dore CJ, Refsum H, Kooner JS. Investigation of          |
| 351 |     | relationship between reduced, oxidized, and protein-bound homocysteine and vascular       |
| 352 |     | endothelial function in healthy human subjects. Circ Res 2001; Jul 20;89(2):187-92.       |
| 353 | 13. | Chambers JC, Obeid OA, Kooner JS. Physiological increments in plasma homocysteine         |
| 354 |     | induce vascular endothelial dysfunction in normal human subjects. Arterioscler Thromb     |
| 355 |     | Vasc Biol 1999 Dec;19(12):2922-7.   |
| 356 | 14. | Sassi S, Cosmi B, Palareti G, Legnani C, Grossi G, Musolesi S, et al. Influence of age,   |
| 357 |     | sex and vitamin status on fasting and post-methionine load plasma homocysteine levels.    |
| 358 |     | Haematologica 2002 Sep;87(9):957-64.  |
| 359 | 15. | De JR, Griffioen PH, van ZB, Brouns RM, Visser W, Lindemans J. Evaluation of a            |
| 360 |     | shorter methionine loading test. Clin Chem Lab Med 2004;42(9):1027-31.                    |
| 361 | 16. | Kalhan SC, LL Gruca, PS Parimi, A O'Brien, L Dierker, and E Burkett. Serine               |
| 362 |     | metabolism in human pregnancy. Am J Physiol Endocrinol Metab 2003; 284: E733-E740.        |
| 363 | 17. | Garcia AJ, Apitz-Castro R. Plasma total homocysteine quantification: an improvement of    |
| 364 |     | the classical high-performance liquid chromatographic method with fluorescence            |

- 365 detection of the thiol-SBD derivatives. *J Chromatogr B Analyt Technol Biomed Life Sci*366 2002; **779**: 359–63.
- 18. Lammarre SG, L MacMillan, GP Morrow, E Randell, T Pongnopparat, M Brosnan et al.
  An isotope-dilution, GC-MS assay for formate and its application to human and animal
  metabolism. *Amino acids* 2014; 46: 1885-1891.
- Kelleher BP, Broin SD. Microbiological assay for vitamin B12 performed in 96-well
   microtitre plates. *J Clin Pathol* 1991; 44: 592-5.13.
- Tamura T, Freeberg LE, Cornwell PE. Inhibition by EDTA of Growth of Lactobacillus
  casei in the folate microbiologic assay and its reversal by added manganese or Iron. *Clin*.

374 *Chem* 1990; 36: 11.

- Bergstrom J, Furst P, Vinnars E. Effect of a test meal, without and with protein, on
  muscle and plasma free amino acids. *Clinical Science* 1990; **79**:331-337.
- 22. Rao S, Yajnik CS, Kanade , Fall CH, Margetts BM, Jackson AA, Shier R et al. Intake of
- 378 micronutrient-rich foods in rural Indian mothers is associated with the size of their babies
- at birth: Pune Maternal Nutrition Study. *J Nutr* 2001;**131**: 1217-24.
- 380 23. Hermann w, H Schorr, R Obeid, and J geisel. Vitamin B-12 status, particularly

381 holotranscobalamin II and methylmalonic acid concentrations, and

- 382 hyperhomocysteinemia in vegetarians. *Am J Clin Nutr*2003;**78**:131-136.
- Antony AC. Vegetarianism and B-12 (cobalamin) deficiency. *Am J Clin Nutr* 2003; **78**:3-6.
- 25. Pawlak R, SE lester and T Babatunde. The prevalence of cobalamin deficiency among
- vegetarians assessed by serum vitamin B12: a review of literature. *Eur J Clin Nutr* 2014;
- **387 68:541-548**.

| 388 | 26. | Quinlivan EP, and JF Gregory. Effect of food fortification on folic acid intake in the   |
|-----|-----|--|
| 389 |     | United States. Am J Clin Nutr 2003; 77:221-225.  |
| 390 | 27. | Gaine PC, MA Pikosky, WF Martin, DR Bolster, CM maresh, and NR rodriguez. Level          |
| 391 |     | of dietary protein impacts whole body protein turnover in trained males at rest.         |
| 392 |     | Metabolism 2006; <b>55</b> : 501-507.  |
| 393 | 28. | Lariviere F, DA Kupranycz, J-L Chiasson, and J Hoffer. Plasma leucine kinetics and       |
| 394 |     | urinary nitrogen excretion in intensively treated diabetes mellitus Am J Physiol 1992;   |
| 395 |     | 263: E173-E179.  |
| 396 | 29. | Sketcher RD, and WP James. Branched-chain amino acid oxidation in relation to            |
| 397 |     | catabolic enzyme activities in rats given a protein-free diet at different stages of     |
| 398 |     | development. British Journal of Nutrition1974; 32: 615-623.                              |
| 399 | 30. | Adibi S Influence of dietary deprivation on plasma concentration of free amino acids of  |
| 400 |     | man. Journal of Applied Physiol. 1968; 25:52-57.   |
| 401 | 31. | Adibi et al Amino acid levels in plasma, liver, and skeletal muscle during protein       |
| 402 |     | deprivation. Am J Physiol. 1973;225:408-414.   |
| 403 | 32. | Nagao et al Adaptational modification of serine and threonine metabolism in the liver to |
| 404 |     | essential amino acid deficiency in rats. Amino acids 2009; 36: 555-562.                  |
| 405 | 33. | Noguchi et al Characterization of dietary protein-dependent anibo acid metabolism by     |
| 406 |     | linking free amino acids with transcriptional profiles through analysis of correlation.  |
| 407 |     | Physiological Genomics 2008; 34: 315-326.  |
| 408 | 34. | Gibson et al Endogenous glycine and tyrosine production is maintained in adults          |
| 409 |     | consuming a marginal-protein diet, Am J Clin Nutr. 2002; 75: 511-518.                    |
| 410 |     |  |

| 412 | 35. | Sheikh K, G Camejo, B Lanne, T Halvarsson, MR Landergren, and ND Oakes.Beyond           |
|-----|-----|---|
| 413 |     | lipids, pharmacological PPARa activation has important effects on amino acid            |
| 414 |     | metabolism as studied in the rat. Am J Physiol Endocrinol Metab 2007;292: E1157-        |
| 415 |     | E1185.  |
| 416 | 36. | Motil KJ, Matthews DE, Bier DM, Burke JF, Munro HN, Young VR. Whole-body                |
| 417 |     | leucine and lysine metabolism: response to dietary protein intake in young men. Am J    |
| 418 |     | Physiol.1981; 240:E712-E721.  |
| 419 | 37. | Kalhan SC, and RW Hanson. Resurgence of serine: an often neglected but indispensable    |
| 420 |     | amino acid. Journal of Biological Chemistry 2012; 287:19786-19791.                      |
| 421 | 38. | Lamarre SG, AM maolloy, SN Reinke, BD Sykes, ME Brosnan, JT Brosnan. Formate            |
| 422 |     | can differentiate between hyperhomocysteinemia due to impaired remethylation and        |
| 423 |     | impaired transsulfuration. Am J Physiol Endocrinol Metab 2012; 302: E61-E67.            |
| 424 | 39. | Refsum H, Yajnik CS, Gadkari M, Schneede J, Vollset SE, Orning L, et al.                |
| 425 |     | Hyperhomocysteinemia and elevated methylmalonic acid indicate a high prevalence of      |
| 426 |     | cobalamin deficiency in Asian Indians. Am J Clin Nutr 2001;74:233-41.                   |
| 427 | 40. | Lu SC. Regulation of hepatic glutathione synthesis: current concepts and controversies. |
| 428 |     | <i>FASEB J</i> ; <b>13</b> : 1169-1183.   |
| 429 | 41. | Finkelstein JD and JJ Martin. Methionine metabolism in mammals. Adaptation to           |
| 430 |     | methionine excess. Journal of Biological Chemistry 1986;261: 1582-1587.                 |
| 431 | 42. | Selhub J and J W Miller. The pathogenesis of homocysteinemia: interruption of the       |
| 432 |     | coordinate regulation by s-adenosylmethionine of the remethylation and transsulfuration |
| 433 |     | of homocysteine. Am J Clin Nutr 1992; 55: 131-138                                       |

- 434 43. Grillo MA and S Colombatto. S-adenosylmethionine and its products. *Amino acids* 2008;
- : 187-193.

438 Figure 1: One-carbon metabolism in-vivo in man, described in detail in the text.

439

- 440 BHMT Betaine-homocysteine S-methyltransferase
- 441 CBS Cystathionine-β-synthase
- 442 GNMT Glycine N-methyltransferase
- 443 MTHFR Methylenetetrahydrofolate reductase
- 444 MS Methionine Synthase
- 445 R Methyl acceptor
- 446 R-CH3 Methylated compound
- 447 SAH S-adenosylhomocysteine
- 448 SAM *S*-adenosylmethionine
- 449 THF Tetrahydrofolate

- 451 Figure 2: Plasma methionine, homocysteine, cysteine and glutathione response to
- 452 methionine load. After an overnight fast each subject received methionine 50 mg/kg mixed
- 453 with orange juice. All data are shown in µmoles/l and represent 50th (25<sup>th</sup> -75<sup>th</sup>) percentiles.
- 454 Squares: American subjects; diamonds: Indian subjects
- 455 Error bars represents 25<sup>th</sup> and 75<sup>th</sup> percentiles.
- 456 Differences between the two groups were tested using Mann-Whitney test .
- 457 \*p<0.05, \*\*p<0.01, \*\*\*p<0.001
- 458 Within group differences from basal were determined using Wilcoxon test.
- 459 Difference in US data shown by # sign. Difference in Indian data shown by + sign.
- 460 **#p<0.05, ##p<0.01, ###p<0.001**
- 461 +**p**<**0.05**, ++**p**<**0.01**, +++**p**<**0.001**
- 462

- Figure 3: Plasma glycine, serine, alanine, taurine, leucine and phenylalanine response to
  methionine load with orange juice. After an overnight fast each subject received
  methionine 50 mg/kg mixed with orange juice. Data shown are μmoles/l and represent 50<sup>th</sup>
  (25<sup>th</sup>-75<sup>th</sup>) percentiles. Squares: American subjects; diamonds: Indian subjects. Error bars
  represents 25<sup>th</sup> and 75<sup>th</sup> percentiles.
- 468 Differences between the two groups were tested using Mann-Whitney test :\*p<0.05, 469 \*\*p<0.01, \*\*\*p<0.001
- 470 Within group differences from basal were determined using Wilcoxon test.
- 471 Difference in US data shown by # sign. Difference in Indian data shown by + sign.
- 472 #**p**<**0.05**, ##**p**<**0.01**, ###**p**<**0.001**
- 473 +**p**<**0.05**, ++**p**<**0.01**, +++**p**<**0.001**
- 474

Figure 4: Correlation between plasma concentration of B6 (Pyridoxal phosphate), and homocysteine in the Indian subjects before (0hr, open) and after (5hr, filled) methionine load. Spearman's rank correlation coefficient: 0 hour r = -0.68 (P<0.05) and 5 hour r = -0.78 (P<0.01).

479

|                          | Indians              | US                   | Р       |
|--------------------------|----------------------|----------------------|---------|
|                          | n=10                 | n=13                 |         |
| Age-y                    | 30.2 (24.0, 34.1)    | 27.0 (25.0, 30.0)    | ns      |
| Height-cm                | 152.0 (147.7, 160.8) | 171.2 (163.5, 175.3) | 0.001   |
| Weight-kg                | 52.6 (45.2, 56.6)    | 71.8 (58.0, 89.1)    | 0.001   |
| BMI-kg/m <sup>2</sup>    | 22.0 (18.7, 25.7)    | 23.3 (20.6, 30.0)    | ns      |
| Vitamin B12-pmoles/l     | 130.5 (104.1,197.1)  | 308.0 (266.0,588.0)  | < 0.001 |
| Folate- nmoles/l         | 19.5 (15.6,21.3)     | 30.0 (27.6,40.7)     | < 0.001 |
| Vitamin B6- nmoles/l     | 38.4 (34.4, 50.3)    | 114.0 (58.2, 165.4)  | 0.001   |
| Vitamin B6 PLP- nmoles/l | 8.9 (8.2, 10.3)      | 13.1 (10.7, 18.8)    | ns      |
| CRP- microg /dl          | 121.7 (47.2, 261.0)  | 165.1 (76.8, 269.5)  | ns      |
| Homocysteine-µmol/l      | 20.4 (16.4, 24.4)    | 7.9 (6.8, 8.9)       | <0.001  |
| Cysteine-µmol/l          | 332.2 (314.6, 346.8) | 371.6 (328.1, 419.3) | ns      |
| Glutathione-µmol/l       | 3.8 (2.2, 3.9)       | 6.5 (5.4, 7.5)       | <0.001  |
|                          |                      |                      |         |

Table 1: Demographic, nutritional and biochemical characteristics of study subjects

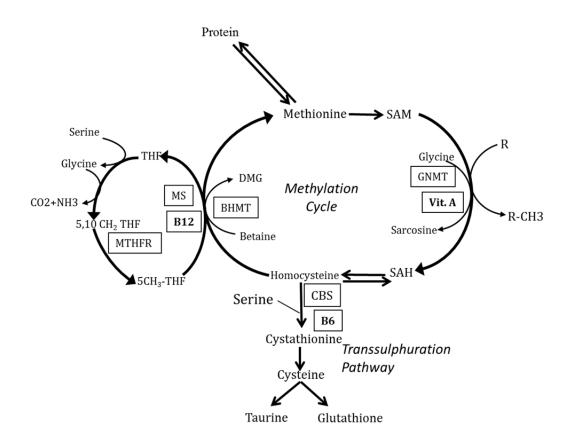
Values are  $50^{\text{th}} (25^{\text{th}} - 75^{\text{th}})$  percentiles. P value by Mann-Whitney test

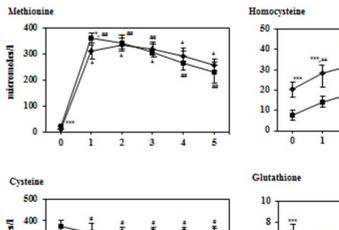
Reference values: B12 > 150 pmoles/l, Folate >7nmoles/l, B6 > 21.2 nmoles/l and tHcy >15 $\mu$ mol/l

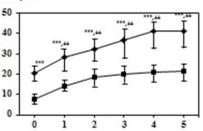
|                   | Indians             | US                     |
|-------------------|---------------------|------------------------|
| Glutamate         | 33.0 (28.5, 43.5)   | 30.0 (21.5, 38.0)      |
| Asparagine        | 46.5 (25.7, 51.2)   | 51.0 (42.5, 63.0)      |
| Serine            | 121.5 (90.7,146.2)  | 107.0 (87.0,123.0)     |
| Glutamine         | 532.0 (380.5,609.2) | 513.0 (477.0,614.5)    |
| Glycine           | 259.0 (210.2,348.0) | 208.0 (180.5,238.5)    |
| Threonine         | 126.5 (83.0,148.2)  | 130.0 (107.5,185.0)    |
| Histidine         | 78.0 (57.0,79.0)    | 98.0 (86.5,111.0)***   |
| Alanine           | 350.0 (287.2,396.5) | 362.0 (301.0,400.0)    |
| Taurine           | 33.5 (26.5,41.2)    | 43.0 (28.5,48.5)       |
| Tyrosine          | 55.5(46.5,63.7)     | 62.0 (45.0,74.0)       |
| Aminobutyric acid | 13.0 (11.0,16.5)    | 22.0 (18.5,26.5)***    |
| Arginine          | 86.0 (62.5,106.5)   | 80.0 (68.5,105.0)      |
| Methionine        | 12.5 (11.2, 14.7)   | 24.0 (19.5, 27.5)***   |
| Valine            | 165.0 (149.2,191.2) | 224.0 (194.0,242.5)*** |
| Tryptophan        | 34.5 (31.0,39.7)    | 51.0 (46.0,60.0)***    |
| Phenylalanine     | 50.0 (43.7,53.0)    | 60.0 (55.5,63.5)**     |
| Isoleucine        | 47.5 (40.0,57.0)    | 58.0 (55.0,67.5)*      |
| Leucine           | 90.5 (80.7,102.2)   | 107.0 (99.0,115.5)**   |
| Ornithine         | 52.5 (37.5,58.7)    | 60.0 (48.5,81.0)       |
| Lysine            | 107.5 (71.7,115.0)  | 201.0 (139.5,277.5)*** |

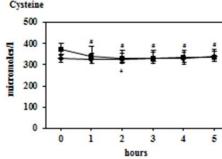
Table 2: Plasma amino acids values (micromoles/l) during fasting

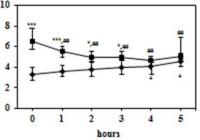
Values are  $50^{\text{th}} (25^{\text{th}} - 75^{\text{th}})$  percentiles .Statistically different compared with the Indian subjects using Mann-Whitney U test \* P<0.05, \*\* P<0.01, \*\*\* P<0.001

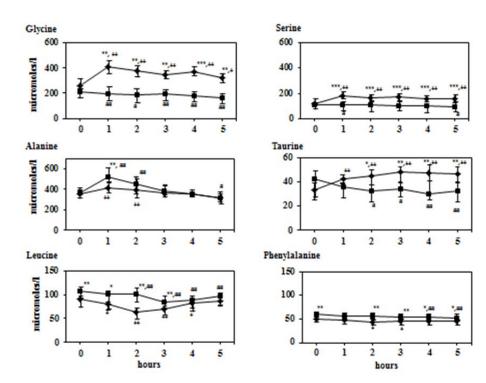


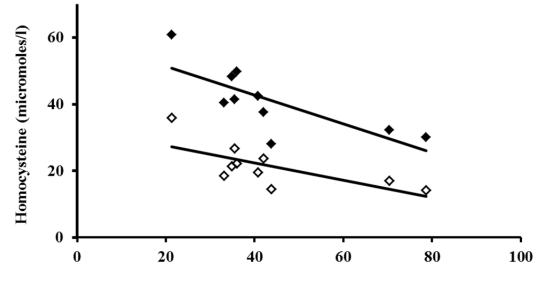












B6 pyridoxal phosphate (nmoles/l)